

REVIEW

Open Access



# Microsponges: a promising frontier for prolonged release-current perspectives and patents

Srinatha N<sup>1\*</sup> , Sowjanya Battu<sup>1</sup> and Vishwanath B. A<sup>1</sup>

## Abstract

**Background** Microsponges are one of the advanced drug delivery systems that facilitates precise and controlled release of active ingredients that are suitable for topical and oral use. These porous microspheres are typically sized between 5 and 300  $\mu\text{m}$ , offer benefits including controlled release, stability, and minimized side effects. Manufacturing techniques like quasi-emulsion solvent diffusion and liquid–liquid suspension polymerization are usually employed to prepare microsponges, although various challenges arise from the use of potentially hazardous organic solvents.

**Main body** Microsponges possess distinct traits such as extended drug release, formulation flexibility, and high drug loading capacity. Entrapment of drugs requires considerations of solubility, stability, and miscibility, while evaluation methods encompass production yield and particle size analysis. Their applications range from dermatological to biopharmaceutical delivery, with diverse products utilizing this technology. Ongoing innovations about microsponges are evident in patents concerning medical dressings and hyaluronic acid delivery systems.

**Conclusion** Microsponges present a promising avenue in drug delivery, despite many challenges. Current review addresses on limitations and diverse products highlighting commercial viability. Patent activity signifies continued interest, suggesting significant potential for enhancing patient care.

**Keywords** Microsponges, Oral administration, Controlled release, Quasi-emulsion solvent diffusion, Topical delivery, Target release

## 1 Background

Innovative drug delivery technology is rapidly evolving and microsponges are at the forefront in innovative pharmaceutical technology. The technology associated with microsphere drug delivery holds immense potential in realizing the objective of precise and regulated drug administration at specific sites. As a result, researchers have given it a lot of attention [1]. Microsponges are

non-collapsible, strongly cross-linked, porous microspheres made of polymeric materials with size ranging between 5 and 300  $\mu\text{m}$  diameter. They possess the ability to load a wide range of active ingredients such as essential oils, antimicrobial agents, fragrances, sunscreens, anti-inflammatory compounds, and antifungal agents. Lately, the use of them for oral administration was also been studied [2, 3, 5]. When used as topical drug carriers, microsponges offer a steady and extended rate of drug release, minimizing discomforts and side effects without compromising the therapeutic efficacy [4]. The purpose of microsponges is to effectively administer a pharmaceutically active substance at the lowest possible dose as well as to improve the formulation stability, elegance, and flexibility, reduce side effects and alter the

\*Correspondence:

Srinatha N  
srinathan304@gmail.com

<sup>1</sup> Department of Pharmaceutics, Aditya Bangalore Institute of Pharmacy Education and Research, #12, Kogilu Main Road, Maruthi Nagar, Behind Annapoorneshwari Temple, Yelankha, Bangalore, Karnataka 560064, India



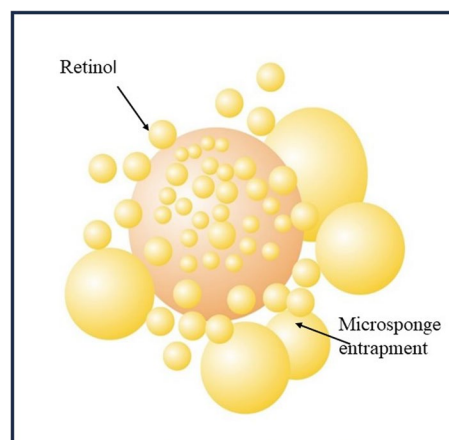
© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

drug release profile in controlled or sustained manner [6]. There are several ways to produce the topical drug formulations using microsphere delivery mechanism, including lotion, gel, or cream. When the microsphere formulation is applied topically, the microsphere delivery system (MDS) releases its active ingredients in response to different stimuli, viz., friction, pH, and temperature at certain timings [7]. Excessive accumulation of drug in the dermis and epidermis may be avoided using microsphere system. As a result, it can considerably decrease irritability without compromising in producing therapeutic activity [6, 8]. Because of their sponge-like structure, microspheres offer special features like easy dissolving and quick compressing of materials. They provide better patient compliance, stable, nontoxic, non-allergic, and non-mutagenic while they offer very few adverse effects unlike other drug delivery systems [2]. Dermal delivery of drug is a broad area of use for microspheres but they are also used for oral administration, advances in bone and tissue engineering, illness detection, and RNAi (ribonucleic acid interference) silencing. The rapid progression of drug delivery technology is further propelled by the emergence of innovative categories of pharmaceuticals and biopharmaceuticals, encompassing proteins, peptides, and nucleic acid-based therapeutics. Consequently, microsphere drug delivery systems represent a burgeoning field that demands thorough exploration [9]. Like enhancement of solubility, precision in targeted organ action, augmented drug stability, targeted medication administration, controlled dispensation of drugs, controlled Release of drugs, dermal drug delivery, oral administration of drugs, advancements in engineering of bone tissue, advances in cardiovascular engineering, rebuilding the vascular walls [10]. Figure 1 shows the structure of microspheres.

## 2 Main text

### 2.1 Characteristics of microspheres [1, 11]

1. Majority of components and vehicles can be used to formulate microspheres.
2. Microspheres exhibit complete miscibility with a small quantity of nonpolar solvent.
3. Microsphere formulations remain stable over the pH range between 1 and 11.
4. Microspheres are stable at temperatures as high as 130° C.
5. Microspheres demonstrate stability when exposed to the catalyst and within the environment of polymerization.
6. Microspheres are self-sterilizing as they possess pores of 0.25  $\mu\text{m}$ , which do not allow bacteria to permeate into them.



**Fig. 1** Structure of microsphere

7. Microsphere compositions can be economical and free flowing.
8. Up to 50–60% of microsphere formulations exhibit substantial entrapment.
9. They are flexible to formulate.
10. It offers prolonged release of drug for up to 12 h.

### 2.2 Advantages of microsphere delivery system [9, 10]

1. Microspheres can absorb oil at a ratio of up to six times their weight without experiencing desiccation.
2. They offer prolonged drug release for up to 12 h.
3. Enhance the robustness at chemical, physical, and thermal levels.
4. Adaptability to create innovative product shapes.
5. They regulate the drug release.
6. Exhibit, improved patient compliance with less irritability and improved tolerance.
7. Can exhibit site specific and targeted activity.
8. MDS have stability over a pH range 1–11.
9. They are free from harmful effects, non-irritating, non-mutagenic, and non-allergic.
10. Exhibits increased drug stability and high drug loading capacity.
11. Compared to other technologies such as liposomes and microencapsulation, MDS is easy to prepare, has a larger payload, and a wider spectrum of chemical stability.
12. MDS allows the incorporation of immiscible products and increases drug's bioavailability.
13. Compatible with all vehicles and other excipients and the solution is easy to navigate and reasonably priced.

### 2.2.1 Advantages over conventional formulations

Conventional topical formulations are formulated with the aim of targeting outer layers of the skin. Upon application, these formulations gradually release their active constituents that form a concentrated coating that absorbs fast. As a result, dermis and epidermis experience an accumulation or excessive buildup of drug, while the microsponges possess the capability to mitigate this issue by releasing the active ingredient to the skin in a gradual manner. Consequently, the micro-sponge system has the potential to significantly diminish side effects such as irritation, while maintaining its efficacy. Examples of such formulations include MDS of benzoyl peroxide, which has minimum irritation and good efficacy [12].

### 2.2.2 Advantages over ointments

Patient compliance is decreased by the ointment's greasy texture. Ointments are not particularly successful as drug delivery vehicles since these compounds require high concentrations of active ingredients to work, which might lead to irritation and sensitization. Bad odors, uncontrollable evaporation of active ingredients and possible drug-vehicle incompatibilities are disadvantages of topical preparations. However, the micro-sponge system in the epidermis or under the skin's surface prolongs the activity without any irritation or other issues faced by normal ointments [13].

### 2.2.3 Advantages over liposomes and microencapsulation

MDS offers benefits in comparison with alternative approaches such as liposomes and microencapsulation. Microcapsules often lack the ability to regulate the rate of active substance leakage. Once the wall ruptures, the active ingredients enclosed within the microcapsules are promptly released. The drawbacks associated with liposomes encompass reduced drug loading, less heat stability, complex formulation processes, limited chemical stability, and susceptibility to microbial instability. Conversely, the micro-sponge system exhibits robustness, enduring temperatures up to 130 °C, and remains stable within a pH range between 1 and 11, distinguishing it from the aforementioned systems. Additionally, its self-sterilizing nature is attributed to an average pore size of 0.25 µm, preventing the access of pathogens while maintaining compatibility with diverse vehicles and substances. Moreover, it retains its free-flowing characteristic and offers a higher drug load capacity ranging between 50 and 60% [14].

### 2.3 Limitations [11]

1. The process of micro-sponge formulation includes addition of organic solvents which are called porogens and are found to be harmful to the environment and public safety as some of them may be very combustible.
2. There are instances where the residual monomer traces found in micro-sponge formulations are poisonous and dangerous to human health.

### 2.4 Properties of the actives for entrapment into microsponges [15–17]

1. Drugs used for micro-sponge formulation should ideally possess minimal solubility; failing which, the vehicle may degrade the micro-sponge before application.
2. Drug must not react with monomers and must not cause the preparation's viscosity to rise while being formulated.
3. It must maintain stability under conditions of polymerization.
4. Should be miscible with minimum quantity of solvent.
5. Drug ought to keep the micro-sponge's spherical structure intact.
6. In order to eliminate cosmetic defects, the vehicle must be restricted to containing only 10 to 12% w/w of micro-sponge.

### 2.5 Micro-sponge preparation methods

#### 2.5.1 Quasi-emulsion solvent diffusion technique

Microsponges by quasi-emulsion solvent diffusion method is prepared by dissolving the polymer in suitable solvent usually ethanol which forms an inner phase. Subsequently, the drug will be added into the inner phase and the mixture was permitted to dissolve for a duration of 15 min at 35 °C under ultrasonication. In the next step, the outer phase is prepared by dissolving polyvinyl alcohol (PVA) in distilled water at an ambient temperature. Following this, the inner phase is combined with the outer phase at room temperature and the mixture is subjected to continuous stirring for a duration of two hours at 500 rpm. This results in micro-sponge formation, later the preparation is filtered to isolate microsponges. Subsequently, the resultant product is cleaned and dried at 40 °C in an oven [18–20].

Figure 2 illustrates the quasi-emulsion solvent diffusion preparation process.

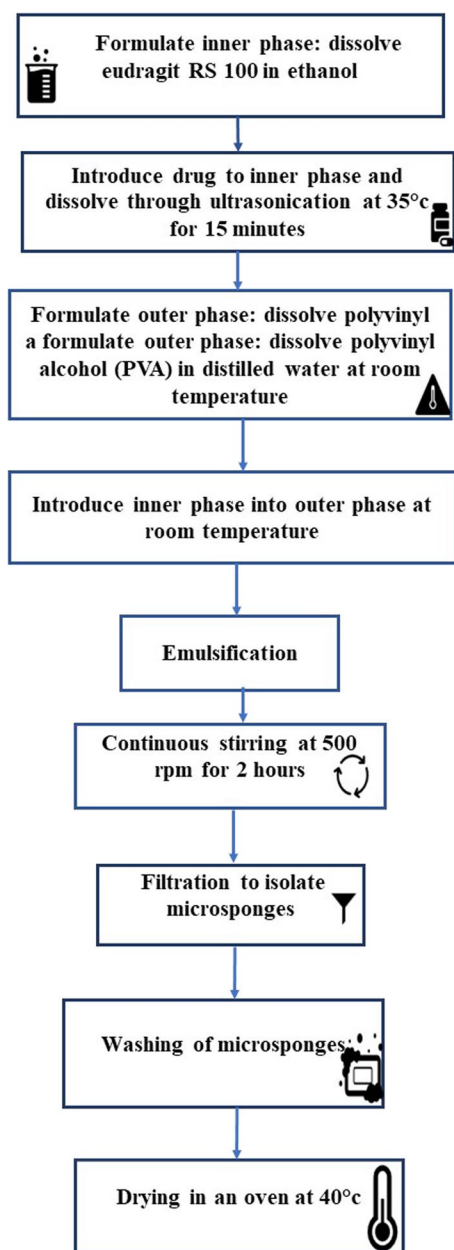


Fig. 2 Quasi-emulsion solvent diffusion preparation method of microsponge

2.5.2 Quasi-emulsion solvent evaporation technique

Quasi-emulsion solvent evaporation technique is one of the very feasible methods to prepare microsponges. Dichloromethane (DCM), ethyl cellulose, and drug were used to create the internal phase and the internal phase is stirred on a magnetic stirrer for 15 min. Subsequently, the internal phase should be cautiously introduced drop by drop into a solution comprising a surfactant and plasticizer in water, which serves as the external phase. Upon the completion of the

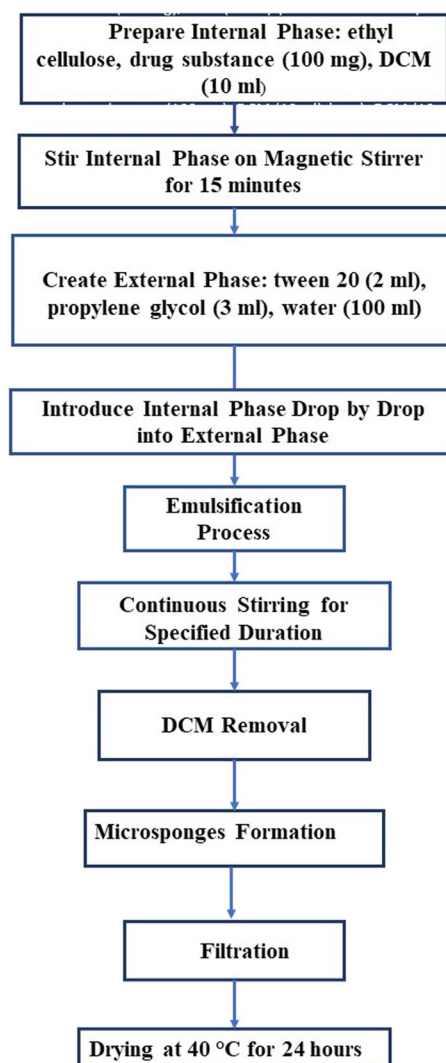


Fig. 3 Quasi-emulsion solvent evaporation preparation method of microsponge

emulsification process, the mixture is subjected to continuous stirring for about 1 h. This results in the elimination of DCM that leads to the formation of microsponges. The suspension formed is thus filtered to obtain microsponges and is dried for 24 hours at 40 °C [21, 22].

Figure 3 illustrates the preparation process for quasi-emulsion solvent evaporation.

2.5.3 Liquid-liquid suspension polymerization

This technique involves the use of monomers, surfactant, and the active ingredient that are dissolved in a suitable solvent. To create a suspension, the above mixture is added with a suspending agent [23]. Once the suspension containing distinct particles of the desired size is

established, polymerization is triggered either through the addition of a catalyst or by raising the temperature, occasionally supplemented with radiation. The polymerization process yields a reservoir-like configuration featuring surface perforations in specific cases, an inert liquid, which is fully miscible with the monomer but immiscible with water, is employed to establish the pore network throughout the polymerization process. Upon completion of the polymerization, the liquid is extracted, resulting in the formation of microsponges that interpenetrate with previously generated microsponges. These microsponges serve as carriers in topical treatments by including a range of active ingredients, including antifungal agents, rubefacients, anti-acne chemicals, and anti-inflammatory agents. A two-stage process is employed when the medication is vulnerable to the conditions of polymerization. Figure 4 illustrates the preparation of suspension polymerization in a liquid–liquid system using a reaction vessel.

An overview of the many steps involved in creating microsponges is provided below:

- Step 1: Selection of monomer and the monomer mixture.
- Step 2: Creation of chain monomers after initiating polymerization.
- Step 3: Ladders are formed by the cross-linking of chain monomers.
- Step 4: Production of spherical particles.

Step 5: Bunches of microspheres are produced when the microspheres agglomerate.

Step 6: Bundles bind together to form microsponges [9, 24].

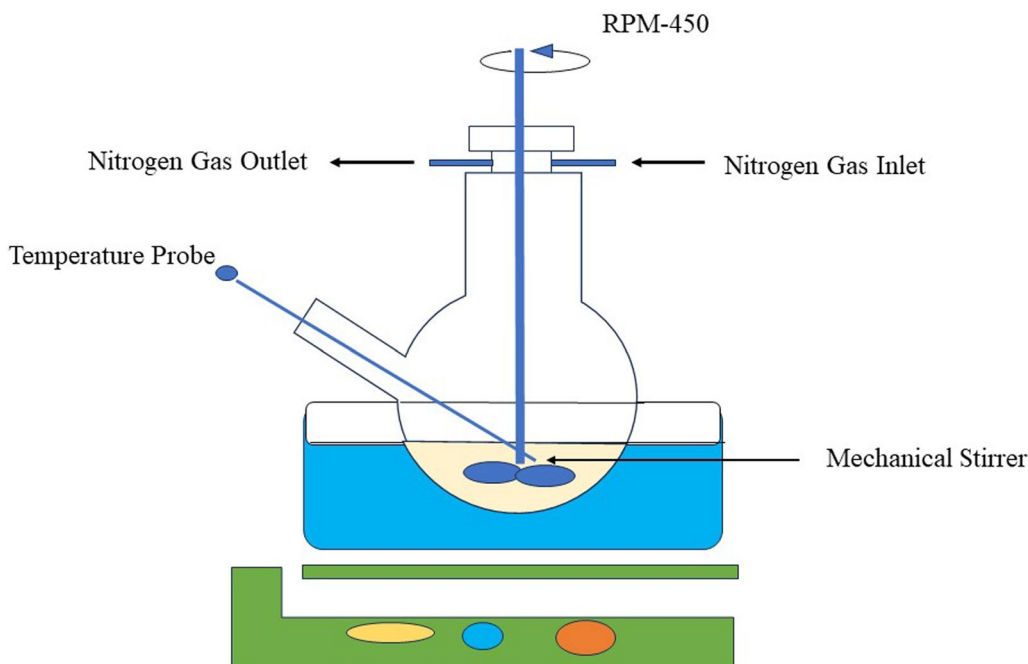
## 2.6 Mechanism of drug release

In reaction to one or more external stimuli, microsponges tend to release drug in a predetermined amount. Figure 5 represents the mechanism of drug release via microsp sponge delivery system.

**Solubility** The rate at which active agents are discharged from microsponges can be triggered by an aqueous media, such as sweat. The release of the active drug can be influenced by factors such as the solubility of the drug in the external medium, the concentration gradient, or the capacity of the microsp sponge network to expand [25].

**Pressure release** When the microsp sponge system is compressed or squeezed, fluid or the active ingredient is released, resupplying the skin with the amount of entrapped active component. The sponge's release and the microsponges' resilience may also have an impact on the amount released [26, 28].

**Temperature change** Temperature can be used to trigger the release of drug from microsponges. At ambient temperature, numerous of the encapsulated active ingredients may exhibit excessive viscosity, hindering their direct flow from the microsponges onto the skin.



**Fig. 4** Preparation of suspension polymerization in a liquid–liquid system using a reaction vessel

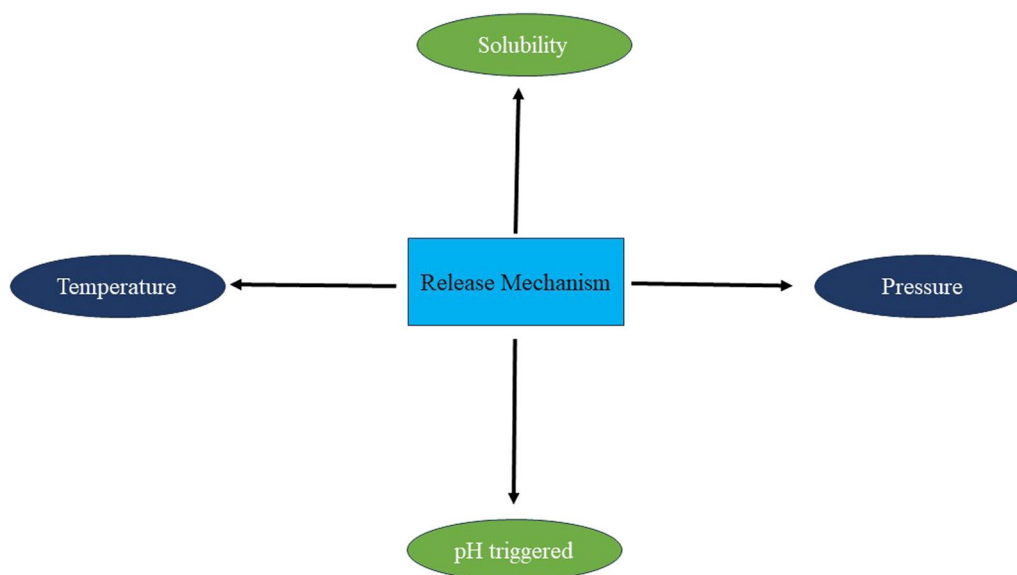


Fig. 5 Mechanism of drug release via microsp sponge delivery system

Raising the temperature of the skin also raises the flow rate, which raises the release [27].

**pH triggered systems** By altering the microsp sponge’s covering or coatings, the microsp sponges can release drug as pH-based released [28].

2.7 Characterization and evaluation of microsp sponges

2.7.1 Production yield

The practical or production yield of the microsp sponges can be calculated using the following formula [29, 30].

$$x = \frac{\text{Actual yield}}{\text{Theoretical yield}} \times 100 \tag{1}$$

2.7.2 Particle size analysis

The particle size of the produced microsp sponges can be analyzed by a particle size analyzer. This instrument enables sample measurement within the range of 0.020 to 2000 mm. The particle size of microsp sponges can also be determined by an optical microscope [31, 32].

2.7.3 Drug content

To estimate the drug content in microsp sponges, 100 mg equivalent microsp sponges are precisely weighed and mixed in 100 milliliters of phosphate buffer solution (PBS) (pH 6.8). The mixture should be filtered through a 0.45-µm membrane filter and the samples are to be analyzed at a suitable wavelength using ultraviolet–visible (UV) spectrophotometer. The drug content can be calculated using the following formula [33].

$$\text{Drug Content(\%)} = \frac{\text{Amount of drug}}{\text{Weighed amount of microsp sponges}} \times 100 \tag{2}$$

2.7.4 Entrapment efficiency

The solvent extraction method can be used to assess the drug entrapment efficiency. Ten mg of precisely weighed microsp sponge particles is dissolved in 5 mL of methanol using a magnetic stirrer for a duration of 20 min. 20 mL of freshly prepared phosphate buffer solution (PBS) must be added and heated to a temperature range of 45–50 °C till the formation of a clear solution. Later, methanol is allowed to evaporate, cooled to 25 °C and filtered. Following appropriate dilutions, the drug’s concentration is measured using UV spectroscopy. To compute drug encapsulation efficiency (DEE%), the following formula can be used [1, 34].

$$\text{DEE\%} = \frac{\text{Actual drug content of microsp sponge}}{\text{Theoretical drug content of microsp sponge}} \times 100 \tag{3}$$

2.7.5 Physical compatibility testing using differential scanning calorimetry (DSC)

This is an essential assessment method for determining any potential physical interactions between the drug and excipients through thermal analysis. Any alteration in the thermogram from that of the pure drug indicates the presence of physical incompatibility. The temperature and enthalpy scale of the DSC are calibrated using indium as the internal standard. Following hermetic

sealing within an aluminum pan, the powder sample of microsponges undergoes a gradual heating process at a rate of 10 °C/min, spanning from 30 to 300 °C, while being subjected to a nitrogen atmosphere flow of 20 ml/min [6, 35].

#### 2.7.6 Scanning electron microscopy (SEM) analysis

The morphology of the microsponges prepared is studied using scanning electron microscopy (SEM) [21, 36].

#### 2.7.7 Chemical compatibility testing using Fourier transform infrared analysis (FTIR)

Using infrared light, the FTIR spectra of the drug-loaded microsponges and the drug alone are examined to search for any chemical interactions. The FTIR analysis is done using a KBr disk standard and scanned within the range of 400 cm<sup>-1</sup> to 4000 cm<sup>-1</sup> [5, 21].

#### 2.7.8 X-ray powder diffraction (XRD)

To comprehend the X-ray diffraction (XRD) pattern of both the pure drug and the optimized formulation, the XRD findings for the samples are acquired using an XRD technique fitted with a nickel filter and copper target. The powder sample is applied uniformly and smoothed out onto metal stages with glass bottoms. The XRD pattern of each sample is obtained with a step increment of 0.10 (2θ) and a dwell time of one second between successive measurements, covering a range from 10 to 500 (2θ) [37, 38].

#### 2.7.9 Stability study

In accordance with “International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use” (ICH) recommendations, various conditions must be set to conduct stability experiments on drug-loaded microsponges. The microsphere formulations are stored under conditions of 40 °C ± 2 °C and 75% ± 5% relative humidity for a duration of three months. Three months later, the microsponges are to be subjected to in vitro drug release studies and physical properties evaluation [2, 38].

#### 2.7.10 Polymer/monomer composition

The drug release from microspheres is controlled by polymer composition, drug loading, and microsphere size. Modifying the drug's partition coefficient between the vehicle and the microsphere system allows for direct influence by the polymer composition of the MDS on the rate of entrapped drug release. A valuable approach for investigating drug release from microsphere systems with diverse polymer compositions involves plotting the cumulative percentage of drug release against time [39].

#### 2.7.11 Resiliency

The rheological properties of the formulation can be modified by changing the concentration of cross-linking agent and polymer composition in order to control its flow characteristics and deformation behavior. On the other hand, one should remember that increasing the concentration of cross-linking agents in order to modify viscosity may lead to the reduction or fluctuations in drug release [40, 41].

#### 2.7.12 In vitro drug release

100 mg equivalent weight of microsponges is weighed accurately and the in vitro drug release studies can be done using USP dissolution testing apparatus type II (USP II). An aliquot of microsphere suspension is administered onto a dialysis membrane (pore size 14,000 Da, diameter 17.5 mm, HI-media) to determine the drug release. The dialysis bags must be fastened using paddles and positioned within dissolution vessels filled with buffer solution. Subsequently, the vessels are subjected to stirring at 50 rpm once the temperature is stabilized to 37 ± 1 °C. Evaluation of drug release into the surrounding solution, attributed to membrane diffusion, is conducted by periodically collecting samples from the solution at specified time intervals. UV visible spectrophotometer can be used to quantify the amount of drug released from microsphere formulation [33, 42].

### 2.8 Different analysis comparing the drugs with various microsphere methods

Listed in Table 1.

### 2.9 Applications of microsphere systems [10]

Porous polymeric microspheres known as microsponges are primarily utilized in topical applications, although recent advancements have expanded their use to oral administration. They offer a diverse range of possibilities for formulators engaged in the creation of pharmaceutical and cosmetic products. Microsponges are formulated to maximize the efficacy of delivering pharmaceutical active ingredients at minimal doses, concurrently elevating stability, mitigating side effects, and modulating drug release dynamics.

1. Microsponges often demonstrate an erratic release profile of active pharmacological ingredients (API). The emergence of the enclosed API within the microsponges is promptly observed upon rupture of the capsule membrane.
2. There exists a payload efficiency ranging between 50 and 60%.

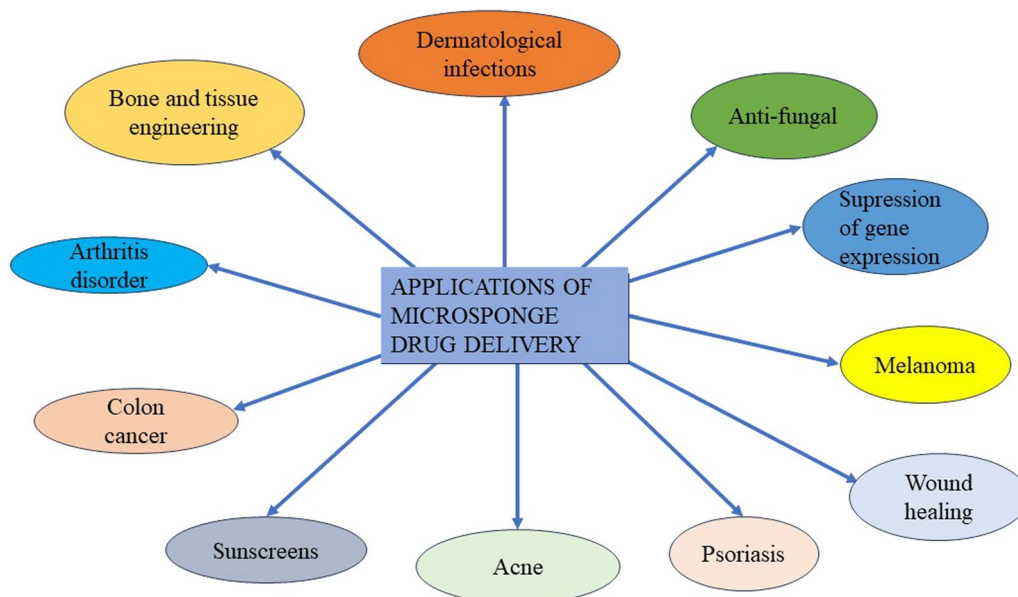
**Table 1** Different analysis comparing the numbers of drugs with microsp sponge methods [43–60]

Drug	Observation	Method of Preparation	References
Honey	The study demonstrated that the optimal honey-loaded microsp sponge formulation possessed favorable antimicrobial and antioxidant properties, suggesting its potential in biomedical applications, including wound healing and tissue regeneration, and warranting further exploration in cosmetics and dermatology	Quasi-emulsion solvent diffusion	[43]
Nebivolol	The swift wound healing observed with the combination of nebigvolol microsp sponge and gel represents a significant advancement in diabetic wound therapy	Oil-in-oil emulsion solvent diffusion	[3]
Diclofenac sodium	The enhanced MDS gel, compared to its commercial counterparts, exhibited minimal skin reactions, favoring regulated diclofenac sodium release into the skin	Double emulsification technique	[4]
Mupirocin	The optimized mupirocin microsp sponge formulation proved skin-stable and non-irritating in the Draize patch test, whereas a microsp sponge-based emulgel demonstrated prolonged efficacy in a rat model of <i>S. aureus</i> -infected surgical wounds	Emulsion solvent diffusion	[44]
Oxybenzone	The oxybenzone-loaded microsp sponge showed significant and improved topical retention of oxybenzone over an extended duration, enhancing the UV protection factor and reducing drug toxicity and irritation compared to marketed formulations	Quasi-emulsion solvent diffusion	[45]
Naringenin	In contrast to the basic naringenin gel, the naringenin-loaded microsp sponge gel exhibited a threefold increase in drug deposition into the skin, offering potential for atopic dermatitis treatment	Quasi-emulsion solvent diffusion	[46]
Erythromycin	Encapsulating erythromycin in a microsp sponge and administering it topically ensures sustained release for 8 h, thereby preventing gastrointestinal inactivation and associated disturbances	Quasi-emulsion solvent diffusion	[47]
Nystatin	The study compared traditional nystatin gel with nystatin-loaded microsp sponge gel, revealing significantly greater drug release in the microsp sponge formulation	Quasi-emulsion solvent diffusion	[37]
Terbinafine hydrochloride	Controlled drug release has been observed to reduce the occurrence of adverse effects, consequently minimizing the need for frequent administration of antifungal gel treatments	Quasi-emulsion solvent diffusion	[48]
Voriconazole	The voriconazole-loaded microsp sponge gel exhibited extended drug release, suggesting its promise as an alternative therapy for fungal infections, potentially surpassing conventional treatments	Quasi-emulsion solvent diffusion	[30]
5-Fluorouracil	In vivo assessment showed that the microsp sponge gel formulation of 5-fluorouracil increased drug deposition by 5.5 times compared to the commercial formulation, significantly reducing skin irritation	Quasi-emulsion solvent diffusion	[49]
Indomethacin	Indomethacin-loaded MDS showed enhanced effectiveness as an analgesic and anti-inflammatory medication compared with conventional indomethacin formulations	Quasi-emulsion solvent diffusion	[19]
Eberconazole nitrate	The microsp sponge gel, carrying the drug, released it in a regulated manner without skin discomfort in rats, and an in vitro study indicated a fourfold increase in drug retention in the stratum corneum layer compared to the commercial eberconazole nitrate cream	Quasi-emulsion solvent diffusion	[50]
Ketoprofen	In comparison with commercial ketoprofen tablets, ketoprofen-loaded MDS showed enhanced bioavailability but exhibited delayed drug release and absorption, thereby improving lag time for drug appearance in plasma and prolonging drug concentration	Quasi-emulsion solvent diffusion	[51]
Paracetamol	The study showed MDS's capacity of MDS to efficiently load high quantities of pharmaceuticals, exhibiting superior loading efficiency compared to alternative microparticle delivery methods	Quasi-emulsion solvent diffusion	[52]
Flurbiprofen	This study found that compression of MDS produces a durable core tablet with prolonged drug release, and employing a pore-plugged approach enables the creation of colon-specific tablets exhibiting zero-order release kinetics	Quasi-emulsion solvent diffusion	[53]
Dicyclomine	Increasing the amount of emulsifying agent correlated with larger microsp sponge particles, while increasing the drug/polymer ratio led to decreased manufacturing yield, smaller particle size with enhanced drug content, and augmenting polymer content improved drug release control	Quasi-emulsion solvent diffusion	[9]



**Table 1** (continued)

Drug	Observation	Method of Preparation	References
Allopurinol	The allopurinol microsponge formulation displayed sustained drug release for 12 h with the requisite entrapment efficiency and buoyancy, and a one-month stability study under accelerated conditions revealed no significant formulation alterations	Quasi-emulsion solvent diffusion	[54]
Famotidine	The famotidine-loaded MDS exhibited a consistent drug -release profile over time	Quasi-emulsion solvent diffusion	[55]
Nitrendipine	Medication characterized by poor water solubility demonstrated a sustained release pattern	Quasi-emulsion solvent diffusion	[56]
Oxiconazole nitrate	The prepared oxiconazole nitrate MDS gel showcased controlled drug release potential, surpassing traditional treatments	Quasi-emulsion solvent diffusion	[6]
Itraconazole	The use of itraconazole in a microsponge drug delivery system has demonstrated controlled release characteristics	Quasi-emulsion solvent diffusion	[7]
Lornoxicam	In vivo comparison showed the superior anti-inflammatory efficacy of microsponge-loaded gel MDS over oral formulations. The beneficial effects extend to rheumatoid arthritis, osteoarthritis, and active lumbar sciatica therapy	Quasi-emulsion solvent diffusion	[57]
Curcumin	Curcumin-loaded microsponge gelatin capsules released 93.2% of curcumin compared with 11.7% from pure curcumin capsules over 8 h, demonstrating superior extended-release capability and greater promise for oral treatment	Quasi-emulsion solvent diffusion	[2]
Etodolac	The creation of the etodolac microsponge altered its release rate, leading to decreased severity of adverse effects	Quasi-emulsion solvent diffusion	[58]
Domperidone	MDS capsules showed a prolonged drug release of 76.38% over 8 h compared to the usual Domstal® formulation, indicating its potential as an alternative therapy for gastroparesis and emesis	Quasi-emulsion solvent diffusion	[59]
Piroxicam	It was found that MDS with a porous, spherical structure could be manufactured, with acceptable physical parameters in the prepared tablets, resulting in a significant enhancement in dissolution rate compared to pure piroxicam tablets	Quasi-emulsion solvent diffusion	[60]



**Fig. 6** Summarized uses of microsponges in various formulations

3. Microsponges, being minute spheres, exhibit the capability to absorb skin secretions.
4. User-friendly and economically viable.

**2.9.1 Pharmaceutical applications [61]**

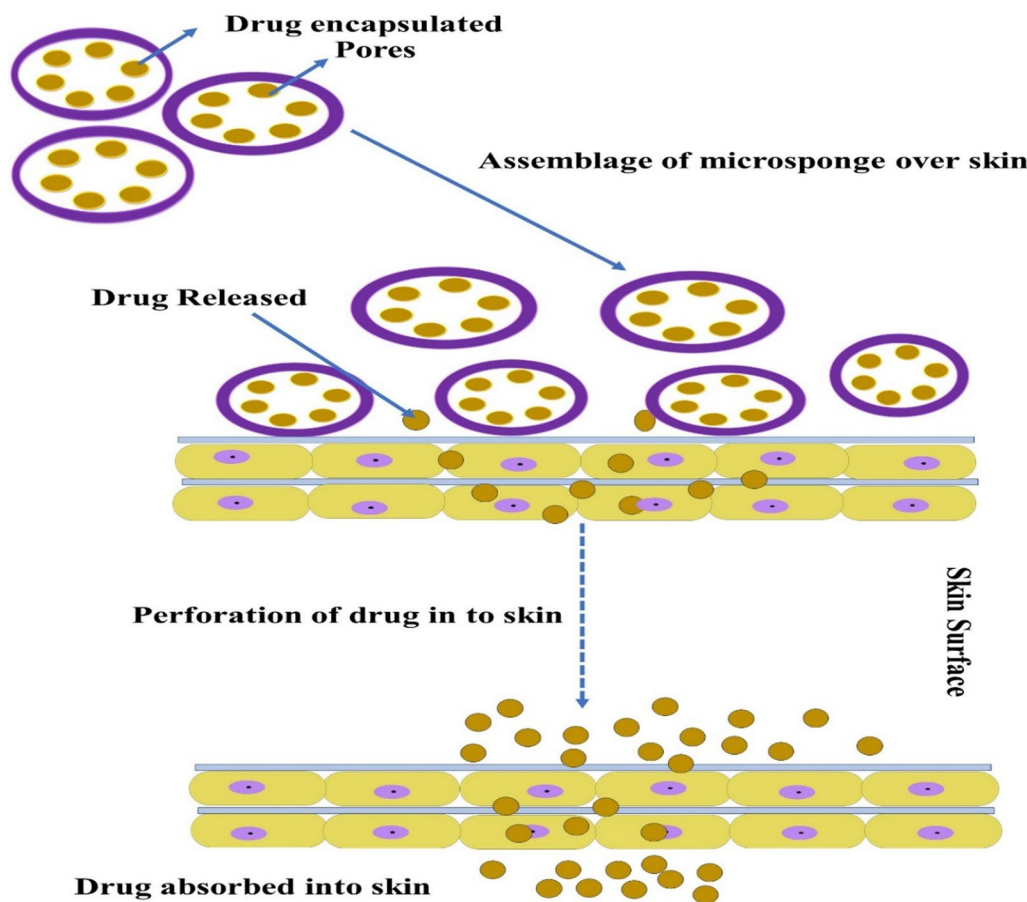
Summarized uses of microsponges in various formulations are listed in Fig. 6.

**2.9.2 Microsponge for topical delivery**

Microsponge systems are fabricated using polymers that demonstrate biologically inert properties. Numerous safety evaluations have confirmed that these polymers exhibit characteristics such as non-toxicity, non-mutagenicity, non-irritation, and non-biodegradability. Because of this, the body is unable to break them down or transform them into other chemicals. These systems, albeit minuscule in size, are too big to fit through the stratum corneum when they are added to topical medicines [17]. Figure 7 shows the mechanism of drug release from dermal microsponges.

Fluocinolone acetonide (FA), a corticosteroid agent, is predominantly employed in dermatology to alleviate skin irritations and improve inflammatory conditions [62]. Acne and athletes’ foot are treated with benzoyl peroxide (BPO). Common adverse effects include skin irritation, which can be lessened while lowering percutaneous absorption by carefully releasing BPO from the microsponge into the skin [63–65].

A research on development of the mupirocin topical microsponges using emulsion solvent diffusion technique has proved to enhance drug deposition in the skin and achieve sustained release. The impact of formulation and procedural parameters including agitation speed and volume of the internal phase, on the physical attributes of microsponges, are explored using an optimized drug/polymer ratio and a 32-factorial design. The enhanced microsponges are integrated into a base prepared with emulgel. Various aspects were examined, including the in vivo antibacterial effectiveness of formulations containing microcin, in vitro drug release, and ex vivo drug deposition. The medication and polymer molecules did not interact with the spherical, porous prepared



**Fig. 7** Mechanisms of drug release from dermal microsponge

microsponges. Preferred physical characteristics were demonstrated by emulgels incorporating microsponges. Drug release assessments conducted with cellulose-based dialysis membranes and drug deposition tests on rat abdominal skin have demonstrated a notable retention of active ingredients within the skin from formulations based on microsponges for up to 24 h. The refined formulations were confirmed to demonstrate stability and skin compatibility test through the Draize patch test. In a murine surgical wound model infected with *S. aureus*, emulgel formulations containing microsponges demonstrated extended effectiveness. The utilization of mupirocin in topical emulgel formulations showed improved retention and stability on the skin, suggesting the delivery system's enhanced efficacy in treating various skin infections, including primary and secondary conditions such as impetigo, atopic dermatitis, and eczema [66].

\*Strong chemotherapeutic medication 5-fluorouracil (5-FU) is used to treat actinic keratosis, a condition caused by chronic sun exposure that results in rigid skin and precancerous cells [67].

### 2.9.3 Microsponge for oral delivery

Microsponges help to maintain the drugs in a protected environment and release the drug under regulated circumstances to the lower gastrointestinal tract [25]. Through the pores of the microsphere system, weakly water-soluble drugs are captured. It has been shown that when drugs are taken orally, the microsphere system can quicken their rate of solubilization. The medicine is effectively reduced to minute particles due to the extremely small pores, which enhances their surface area and accelerates the solubilization process. An additional benefit is that the microsphere system increases the amount of medication absorbed since it takes a lot longer to transit through the small and large intestine [68]. Figure 8 shows the mechanism of drug release from oral microsphere.

A research showed that ketoprofen microsponges were prepared using quasi-emulsion solvent diffusion method and converted into table dosage forms by direct compression technique. This article proved that the plastic deformation of the microsphere structure enhanced compressibility during the physical combination of the medication and polymer, resulting in the production of mechanically robust tablets [69].

Another research used flurbiprofen (FLB) as drug and prepared microsponges using quasi-emulsion solvent diffusion method. Furthermore, FLB was encapsulated within a commercially available Microsphere<sup>®</sup> 5640 device utilizing an entrapment technique. The development of colon-specific formulations involved techniques such as compression coating, pore filling, and tabulation of microsponges with a blend of pectin and hydroxy

propyl methyl cellulose (HPMC). The sponge-like structure of microsponges, allowed for the plastic deformation of the tablet, results in mechanically robust medication distribution tailored to the colon. In vitro analysis demonstrated a modified release pattern of the medication at the eighth hour, aligning with the time of arrival at the proximal colon, particularly in the presence of enzymes within compression-coated tablet formulations designed for colon-specific delivery. Nevertheless, during the eighth hour, the inclusion of enzymes resulted in a notable augmentation in the release of medication from colon-targeted formulations prepared using pore-plugging microsponges [39].

### 2.9.4 Microsponges for bone and tissue engineering as substitute for bone

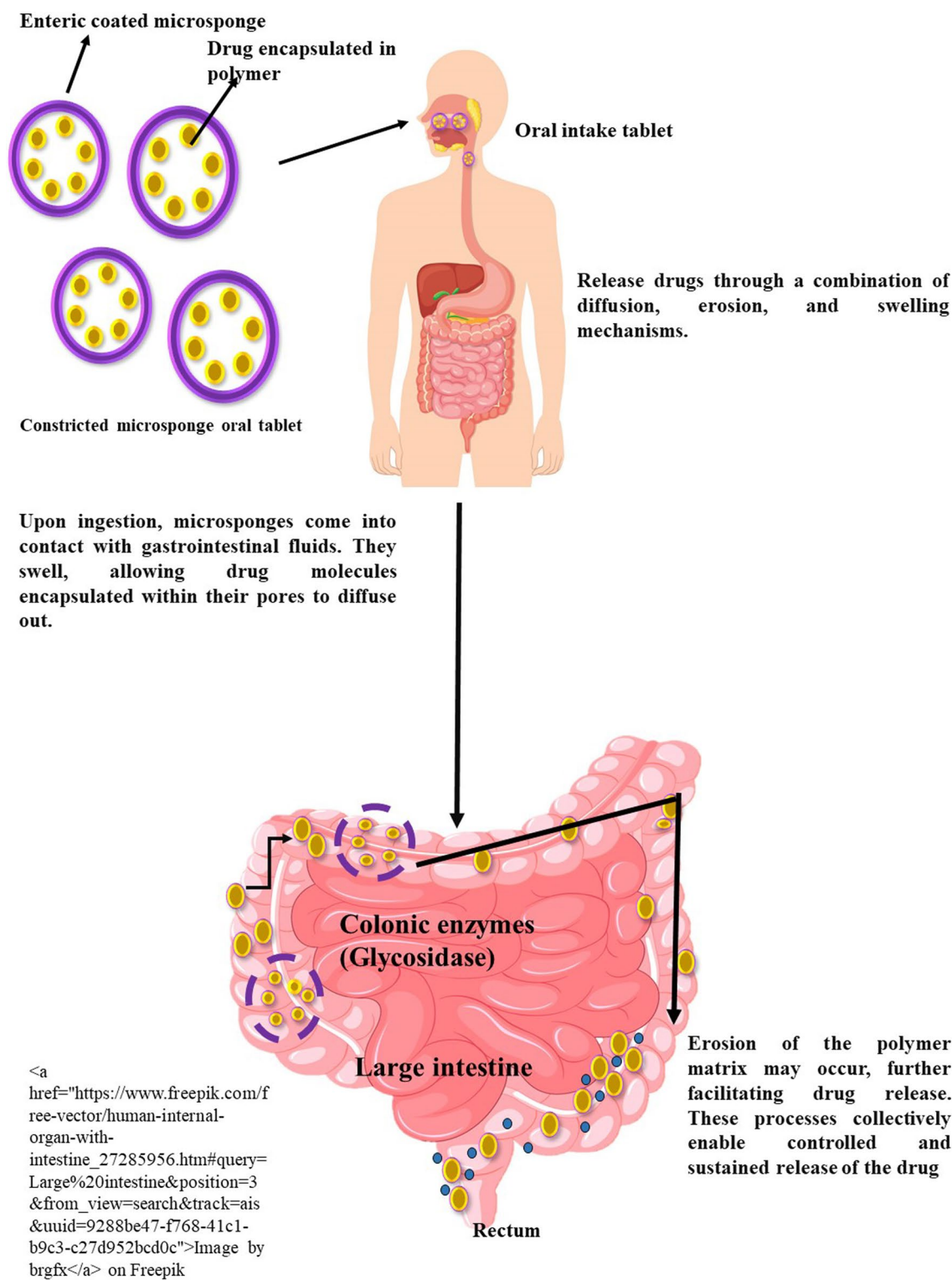
The microsponges were formulated by combining two aqueous dispersions comprising tricalcium phosphate granules, powdered calcium hydroxyapatite, and pre-polymerized polymethyl methacrylate powders with liquid methyl methacrylate monomer. The completed composites seemed porous and worked as microsponges. Utilizing the biodegradable properties of the sponge matrix, a collagen sponge sheet encapsulating basic fibroblast growth factor (bFGF) was administered subcutaneously in mice, demonstrating locally angiogenic activity that varied in accordance with dosage form. The bolus injection of bFGF would never have been able to achieve the substantial increase in blood flow that the collagen microsponges containing bFGF produced in the ischemic hind leg of the mouse. Type I collagen acted as a depot for bFGF, which highlights its significance and its therapeutic use [70–72].

### 2.9.5 Microsponges in oral care cosmetics

By maintaining the release of volatile compounds, microsphere technology offers an exciting prospective application in oral cosmetics, where it can extend the "fresh feel." Tooth pastes and mouth washes can readily absorb microsponges of these volatile substances [73].

### 2.9.6 Microsponges for biopharmaceutical delivery

Biopharmaceutical delivery and tissue engineering both made use of the microsphere delivery technique. The benefits of synthetic poly(lactic-co-glycolic acid) (PLGA-A biodegradable copolymer used in biomedical applications such as drug delivery and tissue engineering) knitted mesh and natural type I collagen were combined to create hybrid 3D scaffolds. For tissue development and cell seeding, collagen microsponges were used, and a mechanically robust PLGA mesh was used as a skeleton. There were three sets of scaffolds: Collagen microsphere could be made in three different ways: sandwich-style (on



**Fig. 8** Mechanisms facilitating the release of drugs from microsponge carriers within an oral drug delivery framework

both sides), semi-thickly (on one side), and thinly (in the crevices between the PLGA mesh) [23].

### 2.9.7 Vascular wall reconstruction with microsp sponge technology

A composite construct was created by integrating a collagen microsp sponge with a biodegradable polymeric scaffold composed of knitted mesh externally reinforced with woven polylactic acid and polyglycolic acid, resulting in the development of a tissue-engineered patch. Tissue-engineered patches devoid of pre-seeding with cells were implanted into the descending aorta of swine (n=5), the main pulmonary artery of pigs (n=8), or the right ventricular outflow tract of canines (n=4). Assessments of the histology and biochemistry were carried after one, two, and six months following the implantation. In each animal, thrombus formation was not observed. Two months post-implantation, histological examination using hematoxylin/eosin staining and immunostaining revealed that all grafts exhibited robust in situ cellularization. The polymerase chain reaction technique, employed to quantify the cell population, detected a significant presence of smooth muscle and endothelial cells two months post-implantation. After six months, the tissue architecture of the patch in the large transplantation model closely resembled that of native tissue, indicating its potential as an innovative surgical material for cardiovascular system restoration [41, 74].

## 2.10 Different analysis showing the difference between different conventional and advanced delivery systems

### 2.10.1 Conventional drug delivery systems

Topical administration

- Traditional methods such as eye drops and ointments are frequently employed in ocular drug delivery owing to their widespread acceptance among patients and the convenience they offer for self-application [75].
- The constrained ocular bioavailability poses a substantial obstacle when utilizing conventional drug delivery systems, thereby affecting the efficiency of drug administration. In this, case microsponges offer a better bioavailability minimizing the limitations observed in conventional systems [76].

### 2.10.2 Advanced drug delivery systems

**2.10.2.1 Novel drug delivery systems (NDDS)** Novel drug delivery systems (NDDS) have been devised to address the shortcomings associated with conventional

formulations, aiming to enhance drug release kinetics, augment drug penetration, and elevate antifungal efficacy.

Research findings indicate that novel drug delivery systems exhibit superior performance compared to conventional formulations in terms of drug release, permeation, and antifungal efficacy, underscoring their advantage in ocular drug delivery [77].

### 2.10.3 Specific differences

**2.10.3.1 Performance** Advanced delivery systems, such as microsponges, exhibit enhanced efficacy regarding drug release, permeation, and therapeutic outcomes when compared to conventional systems.

Microsponges are engineered to optimize drug administration through the augmentation of drug bioavailability and the precise targeting of particular tissues or cellular entities, thereby resulting in enhanced therapeutic efficacy [78].

**2.10.3.2 Antibacterial delivery** Nano-liposomal delivery systems have surfaced as effective transporters for antibacterial agents, providing benefits in encapsulation efficiency, antibacterial mechanisms, and interactions with bioactive compounds [79].

**2.10.3.3 Drug loading and release** Innovative approaches, such as supercritical CO<sub>2</sub> impregnation, have demonstrated elevated drug loadings and extended drug release durations in contrast to conventional soaking methodologies, underscoring the significance of novel techniques in drug delivery systems [80].

### 2.11 Marketed products of microsponges

Listed in Table 2.

### 2.12 Patent information

Patented information is listed in Table 3.

### 2.13 Recent advancements of Microsponges as drug delivery systems

By altering the process of creating microsponges, several innovations have been created, including nanoferrosponges, nanosponges, and microbeads.  $\beta$ -CD nanosponges by using  $\beta$ -CD (beta-cyclodextrin—is a cyclic oligosaccharide commonly used to improve drug solubility and stability in pharmaceutical formulations) is beneficial for trapping of hydrophilic as well as hydrophobic drugs. Several drugs, including flurbiprofen, dexamethasone, doxorubicin, itraconazole, and serum albumin, were administered orally using these methods. In order to make these nanosponges, the  $\beta$ -CD molecule must be cross-linked using biphenyl carbonate. These nanosponge carrier drug delivery systems are particularly

**Table 2** A list of marketed products of microsponges [25, 45, 81–85]

Product name	Manufacturer	Advantages	References
Retinol 15 night cream	Biomedic, sothys	Retinol 15, a night-time therapy cream that utilizes a micro sponge system and pure retinol, consistently improves skin discoloration and visibly reduces fine lines and wrinkles	[25, 81, 82]
EpiQuin micro	Skin Medica Inc	Microsponge® encapsulation of retinol and hydroquinone facilitates their gradual release into the skin, potentially mitigating irritation, and enhancing efficacy over time	[25, 81, 83]
Aramis fragrances	Aramis Inc	An effective antiperspirant spray with 24-h longevity, utilizing microsponge technology to gradually disperse fragrance, facilitated by the absorbent properties of the micro sponge, controlled by environmental factors, offering a free-flowing powder texture	[81, 83, 85]
Ultra guard	Scott paper	Diaper rash protection for babies' skin using a dimethicone-containing micro sponge system	[45, 81, 85]
Retino-A-Micro	Ortho-McNeil Pharmaceutical, Inc	Tretinoin encapsulated within patented porous microspheres at concentrations of 0.1% and 0.04% via microencapsulated drug delivery systems is effective for the topical treatment of acne vulgaris	[45, 81, 82]
Fluorouracil cream, 0.5%	Dermik Laboratories, Inc. Berwyn, PA 19312 USA	A patented porous microsphere (micro sponge) composed of poly(methyl methacrylate-co-glycol dimethacrylate) and dimethicone effectively encapsulated 0.5% fluorouracil within the Carac cream formulation	[25, 81, 82]
Sports cream RS and XS	Embil Pharmaceutical	Microsponge® Delivery System (MDS) encapsulates topical agents with analgesic, anti-inflammatory, and counterirritant properties, tailored for addressing musculoskeletal disorders	[25, 81, 83]
Salicylic acid exfoliating peel 20 and 30	Biophora	Penetrative BHA Exfoliation Treatment with Microsponge Technology, containing 20% to 30% salicylic acid, effectively reduced fine wrinkles, acne, and discoloration. This treatment is especially suited for robust skin types or when expedited results are desired	[81, 83, 85]
Line eliminator dual retinol facial treatment	Avon	Retinol, or vitamin A, in MDS is a lightweight cream that fights wrinkles instantly and over time	[25, 45, 83]
Retinol cream	Biomedic	Microsponge technology enhances retinol retention and, ensures maximal efficacy while minimizing discomfort, thereby supporting the maintenance of healthy mucous membranes, skin, and hair through topical application	[25, 84, 85]
Oil-balancing lotion	Fountain Cosmetics	A lightweight lotion with microsponges absorbs excess oil, providing a matte finish while the skin response complex treats inflammation and tightness in oily, breakout-prone skin	[81, 83]
Lactrex™ 12% moisturizing cream	SDR Pharmaceuticals, Inc., Andover, NJ, S.A. 07821	Lactrex™ incorporates a 12% concentration of lactic acid as a neutral ammonium salt and ammonium lactate, aiding in skin exfoliation. Glycerin and water are included to moisturize and soften dry, flaky, and cracked skin, enhancing its efficacy	[82, 83, 85]

**Table 2** (continued)

Product name	Manufacturer	Advantages	References
Dermalogica oil control lotion	John and Ginger Dermalogica skin care Products	Leveraging moisturizing botanicals, oil-absorbing Microsponge® technology, and an antibacterial skin response complex, this feather-light lotion promotes skin purification, soothing, and hydration	[45, 81, 84]
Micro peel plus	Biomedic	Utilizing Microsponge® technology, the MicroPeel® Plus procedure employs salicylic acid microcrystals to stimulate cellular regeneration, surpassing conventional superficial chemical peels in effectively eliminating dead skin cells without causing harm	[62, 81, 83]
Oil free matte block spf-20	Dermalogica	This sunscreen, utilizing microsponge technology, regulates oil production, provides UV protection, and maintains a matte appearance. Green tea extracts in an oil-free solution reduce inflammation, whereas absorbent microsponges, such as cornstarch and vinyl dimethicone/methicone silica, effectively remove surface oils, enhancing their efficacy	[81, 83, 85]
Glycolic acid moisturizer w/SPF 15	AMCOL Health and Beauty Solutions, Inc. USA	Anti-wrinkle and soothing agent	[82, 85]
NeoBenz®Micro	Intendis Inc. Morristown	It is a keratolytic agent with antibacterial properties	[45, 82]

**Table 3** Patent information on microsponges [86–103]

Patent no.	Publication date	Invention	References
CN107469141B	2020-08-18	A medical dressing containing microsponge and preparation method thereof	[86]
JP6688386B2	2020-04-28	Hyaluronic acid microsponge and method for producing the same	[87]
KR101900387B1	2018-09-20	Microsponges having controlled solubility and improved redissolution property	[88]
US7426776B2	2008-09-23	Nonwoven towel with microsponges	[89]
WO2000072827A2	2000-12-07	Porous drug matrices and methods of manufacture thereof	[90]
US5955109A	1999-09-21	Methods and compositions for topical delivery of retinoic acid	[91]
US5679374A	1997-10-21	Anti-acne composition for the simultaneous treatment of the surface layers and deep layers of the skin, and use thereof	[92]
US5725869A	1998-03-10	Microsphere reservoirs for controlled release application	[93]
US5316774A	1994-05-31	Blocked polymeric particles having internal pore networks for delivering active substances to selected environments	[94]
US5292512A	1994-03-08	Cosmetic or pharmaceutical composition containing microspheres of polymers or of fatty substances filled with at least one active product	[95]
US5145675A	1992-09-08	Two-step methods for the preparation of controlled release formulations	[96]
US5100783	1992-03-31	Weighted microsponge for immobilizing bioactive material	[97]
US4997753A	1991-03-05	Weighted collagen microsponge for immobilizing bioactive material	[98]
CA1288370C	1991-09-03	Weighted collagen microsponge	[99]
WO1986005811	1986-09-10	Weighted microsponge for immobilizing bioactive material	[100]
US4863856A	1989-09-05	Weighted collagen microsponge for immobilizing bioactive materials	[101]
US4861714A	1989-08-29	Weighted collagen microsponge for immobilizing bioactive material	[103]
US4690825A	1987-09-01	Method for delivering an active ingredient by controlled time release utilizing a novel delivery vehicle which can be prepared by a process utilizing the active ingredient as a porogen	[103]

helpful for incorporating cytotoxic medications into targeted drug delivery for malignant cells. Additionally, they noted enhanced ribonucleic acid (RNA) stability and a somewhat successful small interfering RNA (siRNA) encapsulation procedure. This strategy could result in new therapeutic avenues for the delivery of siRNA [26, 104, 105].

#### 2.14 Future prospects

A microsphere is made up of several interconnected voids housed in a non-collapsible framework that can hold a broad range of materials. Today, scientists are focusing more on the delivery of sunscreen, anti-acne, anti-dandruff, and agents that can also be used in the delivery of thermolabile substances like vaccines, proteins, peptides, and deoxyribonucleic acid-based therapeutics. It is also employed in the field of tissue engineering and in controlled drug release systems for medications necessitating extended therapeutic regimens. The outer surface of the sphere is typically porous, allowing substances to flow into and out of it. In these investigations, optimization techniques are used to get the greatest possible result from several formulations. Employs efficient and safe methods for delivering active substance. Moreover, parenteral and pulmonary drug administration using these porous devices has been researched. As microsphere particles find utility as a cell culture medium, their potential extends to stem cell cultivation and cellular regeneration within the organism. Prospective uses of microsphere carrier systems encompass cosmetic applications. Furthermore, the flexibility of the formulation offers advantages across diverse sectors, paving the way for innovative medication delivery systems.

### 3 Conclusion

Microspheres are a revolutionary drug delivery technology with versatile applications. Ranging in size from 5 to 300  $\mu\text{m}$ , these porous microspheres offer controlled release, stability, and reduced side effects. Synthesized through techniques like quasi-emulsion solvent diffusion, they found use in dermatological and oral drug delivery. Advantages include prolonged drug release, adaptability, and high loading capacity, although challenges exist, such as solvent use. Evaluation methods ensure quality, and various preparation techniques contribute to their versatility. Triggered drug release mechanisms enhance effectiveness. Marketed products and patents highlight their commercial viability and ongoing innovations. Microspheres represent a promising frontier in drug delivery, with potential across pharmaceutical and cosmetic domains.

#### Abbreviations

MDS	Microsphere delivery system
DCM	Dichloromethane
SEM	Scanning electron microscope
BPO	Benzoyl peroxide
DNA	Deoxyribonucleic acid
HPMC	Hydroxypropyl methylcellulose
XRD	X-ray powder diffraction
DSC	Differential scanning calorimetry
bFGF	Basic fibroblast growth factor
FA	Fluocinonide acetone
FTIR	Fourier transform infrared analysis
$\beta$ -CD	Beta-cyclodextrin
PLGA	Poly(lactic-co-glycolic acid)
(5-FU)	5-Fluorouracil
PBS	Phosphate-buffered saline
ICH	International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use
RNA	Ribonucleic acid
NDDS	Novel drug delivery systems

#### Acknowledgements

Not applicable.

#### Author contributions

Srinatha N: The majority of the manuscript has been cited from various journals, penned and formatted. Sowjanya Battu: Engaged in the refinement, formatting, and ultimate approval of the preliminary manuscript, and endorsing the final version. Vishwanath B.A: Affirmed the final manuscript.

#### Funding

No funding was obtained for this study.

#### Availability of data and materials

Not applicable.

#### Declarations

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

All the authors declare that they have no competing interests.

Received: 23 February 2024 Accepted: 7 June 2024

Published online: 21 June 2024

#### References

- Design, formulation in-vitro evaluation of herbal microsphere by using. (n.d.). CABI Databases. 4(5):1923–1940. <https://doi.org/10.5555/20153225981>
- Bhatia M, Saini M (2018) Formulation and evaluation of curcumin microspheres for oral and topical drug delivery. *Prog Biomater* 7(3):239–248. <https://doi.org/10.1007/s40204-018-0099-9>
- Pandit AP, Patel SA, Bhanushali VP, Kulkarni VS, Kakad VD (2016) Nebivolol-Loaded Microsphere gel for healing of diabetic wound. *AAPS PharmSciTech* 18(3):846–854. <https://doi.org/10.1208/s12249-016-0574-3>
- Maiti S, Kaity S, Ray S, Sa B (2011) Development and evaluation of xanthan gum-facilitated ethyl cellulose microspheres for controlled percutaneous delivery of diclofenac sodium. *Acta Pharm* 61(3):257–270. <https://doi.org/10.2478/v10007-011-0022-6>



5. Desavathu M, Raghuvveer P, Chunduru M (2017) Design, development and characterization of Valsartan microsponges by quasi emulsion technique and the impact of stirring rate on microsp sponge formation. *J Appl Pharm Sci* 193–198:70128. <https://doi.org/10.7324/japs.2017.70128>
6. Yadav V, Jadhav P, Dombé S, Bodhe A, Salunkhe P (2017) Formulation and evaluation of microsp sponge gel for topical delivery of antifungal drug. *Int J Appl Pharm* 9:30–37. <https://doi.org/10.22159/ijap.2017.9.94.17760>
7. Monika, Dua JS, Prasad D, Hans M, Kumari S (2019) Preparation and characterization of itraconazole microsponges using Eudragit RSPO and study the effect of stirring on the formation of microsponges. *J Drug Deliv Therap* 9:451–458. <https://doi.org/10.22270/jddt.v9i3-s.2879>
8. Kumar JR, Varatharajan R, Muthuraman A (2020) Preparation and evaluation of povidone iodine based microsp sponge for wound healing activity in rats. *J Pharm Sci Res* 12(3):436–442
9. Bhanse Najuka D, Shah C, Shah D (2016) Novel and innovative strategy: microsponges drug delivery system. *Pharma Sci Monit* 7(2):54–73
10. Shafi SK, Duraivel S, Bhowmik D, Kumar KS (2013) Microsp sponge drug delivery system. *Indian J Res Pharm Biotechnol* 1(2):206
11. Khanka PS, Hussain K (2019) Formulation and evaluation of antifungal microsp sponge loaded gel. *Int J Res Eng Sci Manag* 2(12):2581–5792
12. Ahmed A, Makram M, Sayed M, Louis D (2018) An overview of microsp sponge as a novel tool in drug delivery. *MADD* 2(3):1–7. <https://doi.org/10.31031/madd.2018.02.000537>
13. Jyothi KN, Kumar PD, Arshad P, Karthik M, Panneerselvam T (2019) Microsponges: a promising novel drug delivery system. *J Drug Deliv Therap* 9(5-s):188–194. <https://doi.org/10.22270/jddt.v9i5-s.3649>
14. Pradhan SK (2011) Microsponges as the versatile tool for drug delivery system. *Int J Res Pharm Chem* 1(2):243–258
15. Aldawsari H, Badr-Eldin SM (2013) Microsponges as promising vehicle for drug delivery and targeting: preparation, characterization and applications. *Afr J Pharm Pharmacol* 7(17):873–881. <https://doi.org/10.5897/AJPP12.1329>
16. Mantry S, Bagchi A, Das S, Das S (2015) Microsp sponge as a novel strategy of drug delivery system. *Univ J Pharm Sci Res* 1(1):32–38
17. Roy A (2015) Microsp sponge as a novel drug carrier system: a review. *World J Pharm Res* 4(12):680–701
18. Sharma A, Hooda A, Chaudhary H (2016) Formulation and evaluation of topical microsponges of sertaconazole. *World J Pharm Res* 5(11):1444–1461. <https://doi.org/10.20959/wjpr201611-7332>
19. Mahajan Aniruddha G, Jag Tap Leena S, Chaudhari Atul L, Swami Sima P, Mali Prabha R (2011) Formulation and evaluation of microsp sponge drug delivery system using Indomethacin. *IRJP* 2(10):64–69
20. Bhagat VS, Arote SR (2021) Formulation development and in-vitro evaluation of microsp sponge drug delivery system of antifungal drug. *Int J Pure Med Res* 5(3):654–661
21. Redhu S, Pawar N (2021) Development and characterization of microsp sponge gel for topical delivery of oregano oil. *Int J Pharm Sci Res* 12(2):1060–1073. [https://doi.org/10.13040/IJPSR.09758232.12\(2\).1060-73](https://doi.org/10.13040/IJPSR.09758232.12(2).1060-73)
22. Deshmukh K, Poddar SS (2012) Tyrosinase inhibitor-loaded microsp sponge drug delivery system: new approach for hyperpigmentation disorders. *J Microencapsul* 29(6):559–568. <https://doi.org/10.3109/02652048.2012.668955>
23. Vitthal JP, Rajasekaran S (2022) Novel approaches of herbal microsponges design, formulation and characterization: an overview. *Int J Pharm Res Appl* 7(5):1280–1291. <https://doi.org/10.35629/7781-070512801291>
24. Dumbre AK, Banerjee SK, Gadhave MV, Gaikwad DD (2014) Microsp sponge: a novel drug delivery system. <https://www.ajprd.com/index.php/journal/article/view/173>
25. Mansi H (2019) A review on microsp sponge delivery system. *J Drug Deliv Therap* | EBSCOhost. [openurl.ebsco.com](https://openurl.ebsco.com). <https://doi.org/10.22270/jddt.v9i3-s.2938>
26. Jadhav N, Patel V, Mungekar S, Bhamare G, Karpe M, Kadam V (2013) Microsp sponge delivery system: an updated review, current status and future prospects. *J Sci Innov Res* 2(6):1097–1110
27. Thakur R, Kumar S, Gaba P (2020) A review: novel method for microsp sponge drug delivery system. *J Pharm Biol Sci* 15(4):35–44. <https://doi.org/10.9790/3008-1504023544>
28. Lalitha SK, Shankar M, Likhitha D, Dastagiri J, Babu MN (2016) A current view on microsp sponge drug delivery system. *Eur J Mol Biol Biochem* 3(2):88–95
29. Farsana T, Geetha VS, Jumana KK, Mubashira NP (2023) Formulation development and evaluation of antimicrobial drug loaded microsponges for topical drug delivery. *World J Pharm Res*. <https://doi.org/10.20959/wjpr202311-28698>
30. Mohan D (2019) Microsp sponge based drug delivery system of voriconazole for fungal infection: formulation development and In-vitro evaluation. [jddtonline.info](https://doi.org/10.22270/jddt.v9i3.2840). <https://doi.org/10.22270/jddt.v9i3.2840>
31. Dineshmohan S, Gupta VRM (2016) Transdermal delivery of fluconazole microsponges: preparation and in vitro characterization. *J Drug Deliv Therap* 6(6):1334. <https://doi.org/10.22270/jddt.v6i6.1334>
32. Eshwarlal MR, Kishan CV, Krishnarao PV, Motiram CH. Formulation and evaluation of sertaconazole nitrate microsp sponge gel
33. Emerging implementation of drug loaded with microsponges technology and their antifungal activity. *J Pharm Negat Results* 13(S01) (2022). <https://doi.org/10.47750/pnr.2022.13.s01.103>
34. Halder S, Poddar S, Khanam J (2021) Optimization and scale-up methodology in preparing microsp sponge loaded with 5-fluorouracil (5-FU). *Drug Deliv Transl Res*. <https://doi.org/10.21203/rs.3.rs-989826/v1>
35. Rajurkar VG, Tambe AB, Deshmukh VK (2015) Topical anti-inflammatory gels of naproxen entrapped in eudragit based microsp sponge delivery system. *J Adv Chem Eng* 5(2):0122. <https://doi.org/10.4172/2090-4568.1000122>
36. Syed SM (2020) Formulation and evaluation of gel containing fluconazole microsponges. [www.ajprd.com](http://www.ajprd.com). <https://doi.org/10.22270/ajprd.v8i4.753>
37. Bansode AS (2019) Formulation, development and evaluation of Microsp sponge loaded Topical Gel of Nystatin. [jddtonline.info](https://doi.org/10.22270/jddt.v9i2-s.2567). <https://doi.org/10.22270/jddt.v9i2-s.2567>
38. Osmani RAM, Aloorkar NH, Ingale DJ, Kulkarni PK, Hani U, Bhosale RR, Dev DJ (2015) Microsponges based novel drug delivery system for augmented arthritis therapy. *Saudi Pharm J* 23(5):562–572. <https://doi.org/10.1016/j.jsps.2015.02.020>
39. Shinkar DM, Bhamare BS, Saudagar RB (2016) Microsponges. *Asian J Res Pharm Sci* 6(2):77–84. <https://doi.org/10.5958/2231-5659.2016.00011.4>
40. Das A, Chakraborty P, Khatiwara B, Dhakal J, Sarangi S, Singh S, Chakrabarti S (2022) Herbal microsp sponge incorporated sunscreen gel: A novel strategy. *Biomedicine* 42(5):844–850. <https://doi.org/10.51248/v42i5.2016>
41. Pb M, Sg K, Vs H, Yogita S (2015) Recent advances in microsponges drug delivery system. *J Crit Rev* 3(1):2016
42. Wadhwa G, Kumar S, Mittal V, Rao R (2019) Encapsulation of babchi essential oil into microsponges: physicochemical properties, cytotoxic evaluation and anti-microbial activity. *J Food Drug Anal* 27(1):60–70. <https://doi.org/10.1016/j.jfda.2018.07.006>
43. Kothai S, Umamaheswari R (2019) Fabrication and characterisation of honey loaded microsponges. [jddtonline.info](https://doi.org/10.22270/jddt.v9i4.3125). <https://doi.org/10.22270/jddt.v9i4.3125>
44. Amrutiya N, Bajaj A, Madhu MN (2009) Development of microsponges for topical delivery of mupirocin. *AAPS PharmSciTech* 10(2):402–409. <https://doi.org/10.1208/s12249-009-9220-7>
45. Vitthal P, Anuradha S (2020) A review on microsponges drug delivery system. *IJRAR-Int J Res Anal Rev* 7:961–974
46. Nagula RL, Waikar S (2020) Cellulose microsponges based gel of naringenin for atopic dermatitis: design, optimization, in vitro and in vivo investigation. *Int J Biol Macromol* 164:717–725. <https://doi.org/10.1016/j.ijbiomac.2020.07.168>
47. Ravi R, Kumar SS, Parthiban S (2013) Formulation and evaluation of the microsponges gel for an anti-acne agent for the treatment of acne. *Indian J Pharm Sci Res* 3:32–38
48. Thavva VEDAVATHI, Baratam SR (2019) Formulation and evaluation of terbinafine hydrochloride microsp sponge gel. *Int J Appl Pharm* 11(6):78–85
49. Jain SK, Kaur M, Kalyani P, Mehra A, Kaur N, Panchal N (2019) Microsponges enriched gel for enhanced topical delivery of 5-fluorouracil. *J Microencapsul* 36(7):677–691. <https://doi.org/10.1080/02652048.2019.1667447>
50. Bothiraja C, Gholap AD, Shaikh K, Pawar A (2014) Investigation of ethyl cellulose microsp sponge gel for topical delivery of eberconazole nitrate

- for fungal therapy. *Ther Deliv* 5(7):781–794. <https://doi.org/10.4155/tde.14.43>
51. Avoxa-Mediengruppe Deutscher Apotheker GmbH (n.d.) Enhancement of ketoprofen bioavailability by formation of microsp.: Ingenta Connect. [www.ingentaconnect.com](http://www.ingentaconnect.com). <https://doi.org/10.1691/ph2007.1.6016>
  52. Jain V, Singh R (2011) Design and characterization of colon-specific drug delivery system containing paracetamol microsponges. *Arch Pharmacol Res* 34(5):733–740. <https://doi.org/10.1007/s12272-011-0506-4>
  53. Orlu M, Cevher E, Araman A (2006) Design and evaluation of colon specific drug delivery system containing flurbiprofen microsponges. *Int J Pharm* 318(1–2):103–117. <https://doi.org/10.1016/j.ijpharm.2006.03.025>
  54. Patel D, Gohil D, Patel D, Shah H, Patel S, Pandya K, Shah C (2016) Formulation and evaluation of floating microsponges of allopurinol. *Pharma Sci Monit* 7(3):135–154
  55. Charagonda S, Puligilla RD, Ananthula MB, Bakshi V (2016) Formulation and evaluation of famotidine floating microsponges. *Int Res J Pharm* 7(4):62–67
  56. Yang M, De Cui F, You BG, Fan Y, Wang L, Peng Y, Yang H (2003) Preparation of sustained-release nitrendipine microspheres with Eudragit RS and Aerosil using quasi-emulsion solvent diffusion method. *Int J Pharm* 259(1–2):103–113. [https://doi.org/10.1016/s0378-5173\(03\)00209-6](https://doi.org/10.1016/s0378-5173(03)00209-6)
  57. He Y, Majid K, Maqbool M, Hussain T, Yousef AM, Khan IU, Mehmood Y, Aleem A, Arshad MS, Younus A, Nirwan JS, Ghori MU, Rizvi SAA, Shahzad Y (2020) Formulation and characterization of lornoxicam-loaded cellulosic-microsponge gel for possible applications in arthritis. *Saudi Pharm J* 28(8):994–1003. <https://doi.org/10.1016/j.sjps.2020.06.021>
  58. Swetha A, Rao MG, Ramana KV, Basha BN, Reddy VK (2011) Formulation and in vitro evaluation of etodolac entrapped in microsponge based drug delivery system. *Int J Pharm* 1(2):73–80
  59. Osmani RAM, Aloorkar NH, Thaware BU, Kulkarni PK, Moin A, Hani U, Srivastava A, Bhosale RR (2015) Microsponge based drug delivery system for augmented gastroparesis therapy: formulation development and evaluation. *Asian J Pharm Sci* 10(5):442–451. <https://doi.org/10.1016/j.ajps.2015.06.003>
  60. Rajab NA, Jawad MS (2016) Formulation and in vitro evaluation of piroxicam microsponge as a tablet. *Int J Pharm Pharm Sci* 8(2):104–114
  61. Afnan T, Chakraborty P, Chakraborty DD, Chhetri P (2022) Microsponge based drug delivery systems: a critical update on its preparation, dermatological applications, and patent information. *J Chengdu Univ Technol* 26:24
  62. D'Souza JI, More HN (2008) Topical anti-inflammatory gels of fluocinolone acetonide entrapped in eudragit based microsponge delivery system. <https://tjptonline.org/AbstractView.aspx?PID=2008-1-4-101>
  63. Jelvehgari M, Siahi-Shadbad MR, Azarmi S, Martin GP, Nokhodchi A (2006) The microsponge delivery system of benzoyl peroxide: preparation, characterization and release studies. *Int J Pharm* 308(1–2):124–132. <https://doi.org/10.1016/j.ijpharm.2005.11.001>
  64. Nokhodchi A, Jelvehgari M, Siahi MR, Mozafari MR (2007) Factors affecting the morphology of benzoyl peroxide microsponges. *Micron* 38(8):834–840. <https://doi.org/10.1016/j.micron.2007.06.012>
  65. Wester RC, Patel R, Nacht S, Leyden JJ, Melendres J, Maibach HI (1991) Controlled release of benzoyl peroxide from a porous microsphere polymeric system can reduce topical irritancy. *J Am Acad Dermatol* 24(5):720–726. [https://doi.org/10.1016/0190-9622\(91\)70109-f](https://doi.org/10.1016/0190-9622(91)70109-f)
  66. Jain N, Sharma PK, Banik A (2011) Recent advances on microsponge delivery system. *Int J Pharm Sci Rev Res* 8(2):13–23
  67. Mandava SS, Thavva V (2012) Novel approach: microsponge drug delivery system. *Int J Pharm Sci Res* 3(4):967. [https://doi.org/10.13040/IJPSR.0975-8232.3\(4\).967-80](https://doi.org/10.13040/IJPSR.0975-8232.3(4).967-80)
  68. Aloorkar NH, Kulkarni AS, Ingale DJ, Patil RA (2012) Microsponges as innovative drug delivery systems. *Int J Pharm Sci Nanotechnol* 5(1):1597–1606
  69. Thakur I, Sharma N (2021) A review on innovative and novel strategy-floating microsponges. *Zenodo*. <https://doi.org/10.5281/zenodo.4879516>
  70. Kaity S, Maiti S, Ghosh AK, Pal D, Ghosh A, Banerjee S (2010) Microsponges: a novel strategy for drug delivery system. *Agric Policy Pap* 1(3):283. <https://doi.org/10.4103/0110-5558.72416>
  71. Kappor D, Patel MP, Vyas R, Lad C, Tyagi B (2014) A review on microsponge drug delivery system. *J Drug Deliv Therap* 4(5):978. <https://doi.org/10.22270/jddt.v4i5.978>
  72. Hussain H, Juyal D, Dhyani A (2014) Microsponges: an overview. *Int J Drug Deliv Technol* 4(4):58–66
  73. Shah CN, Shah DP (2014) Microsponges: a revolutionary path breaking modified drug delivery of topical drugs. *Int J Pharm Res* 6(2):1–13
  74. Iwai S, Sawa Y, Ichikawa H, Taketani S, Uchimura E, Chen G, Hara M, Miyake J, Matsuda H (2004) Biodegradable polymer with collagen microsponge serves as a new bioengineered cardiovascular prosthesis. *J Thorac Cardiovasc Surg* 128(3):472–479. <https://doi.org/10.1016/j.jtcvs.2004.04.013>
  75. Maurya P, Fatma S, Sharma D, Mishra JN, Kushwaha A (2022) Ocular drug delivery systems: an overview. *IJRDO J Appl Sci* 8(11):26–30. <https://doi.org/10.53555/as.v8i11.5439>
  76. Mehrandish S, Mirzaeei S (2020) A review on ocular novel drug delivery systems of antifungal drugs: functional evaluation and comparison of conventional and novel dosage forms. *Adv Pharm Bull* 11(1):28–38. <https://doi.org/10.34172/apb.2021.003>
  77. Duxfield L, Sultana R, Wang R, Englebretsen V, Deo S, Rupenthal ID, Al-Kassar R (2015) Ocular delivery systems for topical application of anti-infective agents. *Drug Dev Ind Pharm* 42(1):1–11. <https://doi.org/10.3109/03639045.2015.1070171>
  78. Shi P, Cheng Z, Zhao K, Chen Y, Zhang A, Gan W, Zhang Y (2023) Active targeting schemes for nano-drug delivery systems in osteosarcoma therapeutics. *J Nanobiotechnol* 21(1):103. <https://doi.org/10.1186/s12951-023-01826-1>
  79. Yousefi M, Andishmand H, Assadpour E, Barzegar A, Kharazmi MS, Jafari SM (2023) Nanoliposomal delivery systems of natural antibacterial compounds; properties, applications, and recent advances. *Crit Rev Food Sci Nutr*. <https://doi.org/10.1080/10408398.2023.2170318>
  80. Coutinho IT, Maia-Obi LP, Champeau M (2021) Aspirin-loaded polymeric films for drug delivery systems: comparison between soaking and supercritical CO<sub>2</sub> impregnation. *Pharmaceutics* 13(6):824. <https://doi.org/10.3390/pharmaceutics13060824>
  81. Chandramouli Y, Firoz S, Yasmeen BR, Vikram A, Mahitha B, Aruna U (2012) Microsponges: a novel drug delivery system for controlled delivery of topical drugs. *Int J Pharm Res Appl* 2(2):79–86
  82. Tiwari A, Tiwari V, Palaria B, Kumar M, Kaushik D (2022) Microsponges: a breakthrough tool in pharmaceutical research. *Future J Pharm Sci* 8(1):10. <https://doi.org/10.1186/s43094-022-00421-9>
  83. Vishwakarma P, Choudhary R (2019) Microsponges: A novel strategy to control the delivery rate of active agents with reduced skin irritancy. *J Drug Deliv Therap* 9(6-s):238–247. <https://doi.org/10.22270/jddt.v9i6-s.3757>
  84. Shaha V, Jain H, Krishna J, Patel P (2010) Microsponge drug delivery: a review. *Int J Res Pharm Sci* 1(2):212–218
  85. Mahant S, Kumar S, Nanda S, Rao R (2020) Microsponges for dermatological applications: perspectives and challenges. *Asian J Pharm Sci* 15(3):273–291. <https://doi.org/10.1016/j.ajps.2019.05.004>
  86. CN107469141A—A kind of microsponge medical dressing and preparation method thereof—Google Patents. <https://patents.google.com/patent/CN107469141A/en>
  87. JP6688386B2—Hyaluronic acid microsponge and method for producing the same Google Patents. <https://patents.google.com/patent/JP6688386B2/en>
  88. KR101900387B1—Microsponges having controlled solubility and improved redissolution property—Google Patents. <https://patents.google.com/patent/KR101900387B1/en>
  89. Love FS III (n.d.) US7426776B2—Nonwoven towel with microsponges—Google Patents. <https://patents.google.com/patent/US7426776>
  90. Straub J (n.d.) WO2000072827A2—Porous drug matrices and methods of manufacture thereof—Google Patents. <https://patents.google.com/patent/WO2000072827A2/en>
  91. Won R (n.d.) US5955109A—Methods and compositions for topical delivery of retinoic acid—Google Patents. <https://patents.google.com/patent/US5955109A/en>
  92. Fanchon C (n.d.) US5679374A—Anti-acne composition for the simultaneous treatment of the surface layers and deep layers of the skin, and use thereof—Google Patents. <https://patents.google.com/patent/US5679374A/en>

93. Lo RJR, Ltd Z (n.d.) US5725869A—Microsphere reservoirs for controlled release application—Google Patents. <https://patents.google.com/patent/US5725869A/en>
94. Eury RP (n.d.) US5316774A—Blocked polymeric particles having internal pore networks for delivering active substances to selected environments—Google Patents. <https://patents.google.com/patent/US5316774A/en>
95. Schaefer H (n.d.) US5292512A—Cosmetic or pharmaceutical composition containing microspheres of polymers or of fatty substances filled with at least one active product—Google Patents. <https://patents.google.com/patent/US5292512A/en>
96. Won R (n.d.) US5145675A—Two step method for preparation of controlled release formulations—Google Patents. <https://patents.google.com/patent/US5145675A/en>
97. Khule PK, Nitalikar MM, More VV, Gilhotra RM (2019) Microsponge drug delivery: a review. *SGVU J Pharm Res Educ* 4(1):359–365
98. Dean RC Jr. US4997753A—Weighted collagen microsponge for immobilizing bioactive material—Google Patents. <https://patents.google.com/patent/US4997753A/en>
99. Dean RC Jr, Berg RA, Phillips PG, Runstadler PW Jr, Silver FH, Corp V (n.d.) CA1288370C—Weighted collagen microsponge—Google Patents. <https://patents.google.com/patent/CA1288370C/en>
100. Dean RC, Phillips PG, Runstadler PW Jr, Silver FH, Berg RA, Cahn F, Corporation V (n.d.) WO1986005811A1—Weighted microsponge for immobilizing bioactive material—Google Patents. <https://patents.google.com/patent/WO1986005811A1/en>
101. Dean RC Jr, Silver FH, Berg RA, Phillips PG, Runstadler PW Jr, Maffia GJ, Corp V (n.d.) US4863856A—Weighted collagen microsponge for immobilizing bioactive materials—Google Patents. <https://patents.google.com/patent/US4863856A/en>
102. Dean RC Jr, Silver FH, Berg RA, Phillips PG, Runstadler PW Jr, Corp V (n.d.) US4861714A—Weighted collagen microsponge for immobilizing bioactive material—Google Patents. <https://patents.google.com/patent/US4861714>
103. Won R (n.d.) US4690825A—Method for delivering an active ingredient by controlled time release utilizing a novel delivery vehicle which can be prepared by a process utilizing the active ingredient as a porogen—Google Patents. <https://patents.google.com/patent/US4690825A/en>
104. Cavalli R, Trotta F, Tumiatti W (2006) Cyclodextrin-based nanosponges for drug delivery. *J Incl Phenom Macrocycl Chem* 56(1–2):209–213. <https://doi.org/10.1007/s10847-006-9085-2>
105. Valluru R, Ravi G, Bose SP, Damineni S (2019) Microsponges—a comprehensive review: success and challenges. *Indo Am J Pharm Res* 9(7):3056–3067

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.